

**IMMATURE STAGES OF *RHYPARONOTUS ALTARENSIS*  
(OLLIFF) (COLEOPTERA: CURCULIONIDAE: MOLYTINAE),  
WITH COMMENTS ON LARVAL CHARACTERS IN  
ANCHONINI AND MOLYTINAE**

ADRIANA E. MARVALDI\* AND JUAN J. MORRONE\*\*

\*Instituto Argentino de Investigaciones de las Zonas Áridas (IADIZA), C.C. 507,  
5500 Mendoza, Argentina; and

\*\*Museo de Zoología, Facultad de Ciencias, Universidad Nacional Autónoma de  
México (UNAM), Apdo. Postal 70-399, México 04510 D.F., México

*Abstract.*—The last instar larva and pupa of *Rhyparonotus altarensis* (Olliff) (Curculionidae: Molytinae: Anchonini), from Ecuador, are described and illustrated. Larval characters are compared with those of other Molytinae, e.g., Molytini, Hylobiini, and Phrynixini, and their phylogenetic value is discussed.

---

The subfamily Molytinae (recently demoted to a tribe Molytini of Curculioninae, see Kuschel, 1995) resulted from the amalgamation of about 30 tribes and subfamilies by Kuschel (1987), to which Dinomorphini (Thompson, 1992) and Bagoiini (Morrone, 1997) were subsequently added (see Appendix). This cosmopolitan subfamily comprises more than 3,000 species and 300 genera (Thompson, 1992). Although several authors have treated molytines as a distinct taxon, either of tribal (May, 1993; Kuschel, 1995; Morrone & Roig-Juñent, 1995; Marvaldi, 1997) or subfamilial rank (Wibmer & O'Brien, 1986; Kuschel, 1987; Thompson, 1992; May, 1994; Zimmerman, 1994; Morrone, 1997), it is debatable whether or not this highly speciose group really represents a natural taxon. Calder (1989) has pointed out the enormous diversity of anatomical features among some of the taxa included in Kuschel's Molytinae, suggesting that this taxon was not a monophyletic group. May (1994) considered that the larvae of many genera included in Molytinae diverge so far from the typical *Hylobius* that a diagnosis of the group "would serve no useful purpose." Doubts on the monophyly of Molytinae are shared by other authors (Robin Craw, pers. comm.).

The molytine tribe Anchonini is a relatively small group (about 200 species), ranged in the Americas (from the USA to Bolivia), Japan and the Oriental region (Faust, 1892; Morimoto, 1982; O'Brien & Wibmer, 1982; Wibmer & O'Brien, 1986; Voisin, 1992, 1993). Our cursory review of the literature shows that knowledge about the biology and immature stages of this taxon is very scarce. A brief larval description of *Anchonus* Schoenherr, based on larvae of unidentified species from Bahamas and Panama, was provided by Anderson (1952), who also intended a larval definition of the tribe Anchonini. However, little confidence can be given to that tribal diagnosis as it was based on larvae of taxa not currently placed in Anchonini, except those assigned to *Anchonus*. A recently fallen *Croton* sp. tree (Euphorbiaceae) in Ecuador provided one of us (JJM) the opportunity to collect specimens of *Rhyparonotus*

*altarensis* (Olliff). When the bark of the tree was removed, three mature larvae, two pupae, and one adult of this species were discovered.

Our main objective is to describe the immature stages of this anchonine species. Furthermore, we discuss the phylogenetic value of some characters that could help elucidating the placement of the tribe Anchonini, as well as testing the monophyly of the subfamily Molytinae.

The specimens examined for this study have the following collection data: EC-UADOR Napo: 52 km N Tena, 6500 feet, under bark of *Croton* sp., 24-V-95, J. J. Morrone coll., det. by associated adult, 3 mature larvae, 2 pupae. The immature specimens have been deposited in the collection of the Instituto Argentino de Investigaciones de las Zonas Áridas (IADIZA), and the associated adult in the collection of the Museo de Zoología, Facultad de Ciencias (UNAM). Techniques for dissection of larvae and terminology and abbreviations here applied are those used in Marvaldi (1998). Pupae were studied following May (1987, 1994).

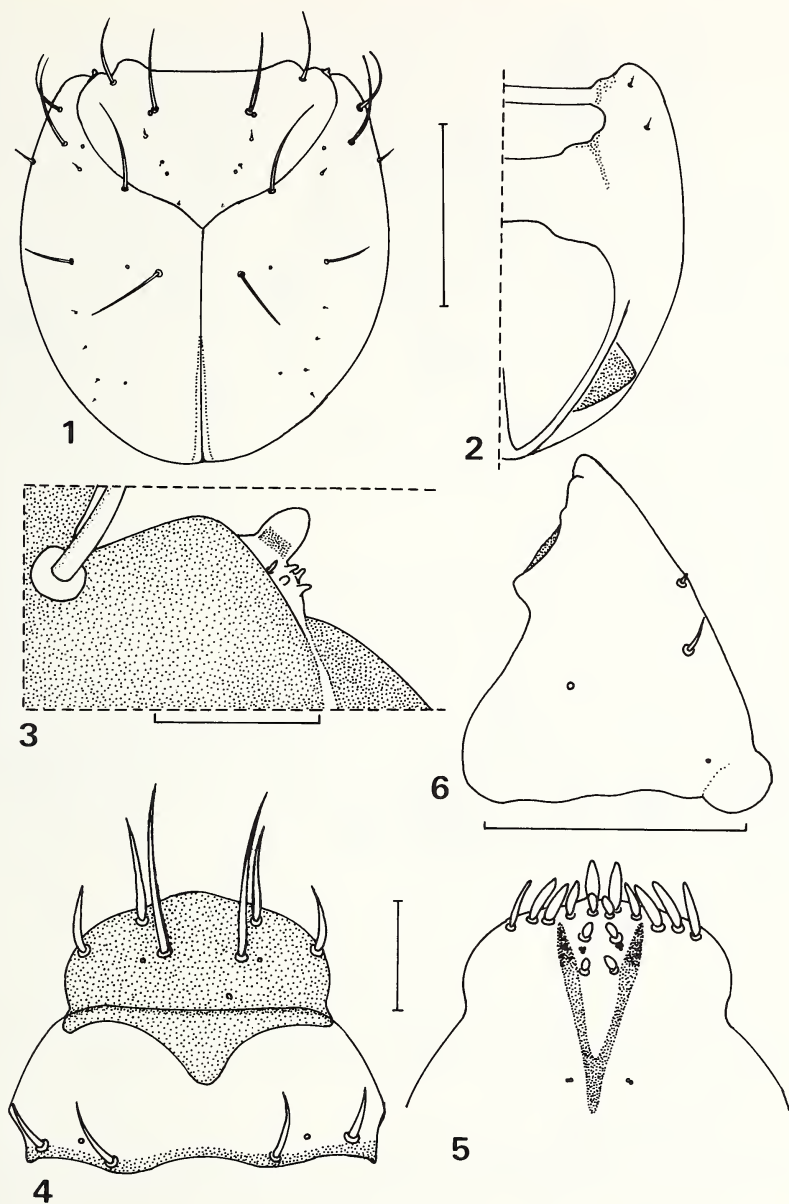
#### IMMATURE STAGES OF *RHYPARONOTUS ALTARENSIS* (OLLIFF)

**Description.** *Mature larva* (Figs. 1–14). Maximum size 13.28 mm × 3.75 mm. Head width 2.20 mm. Body robust, cuticle finely spiculate, setae rather short except for long dorsal and pleural setae on AVII–IX; AVII and AIX transverse, depressed.

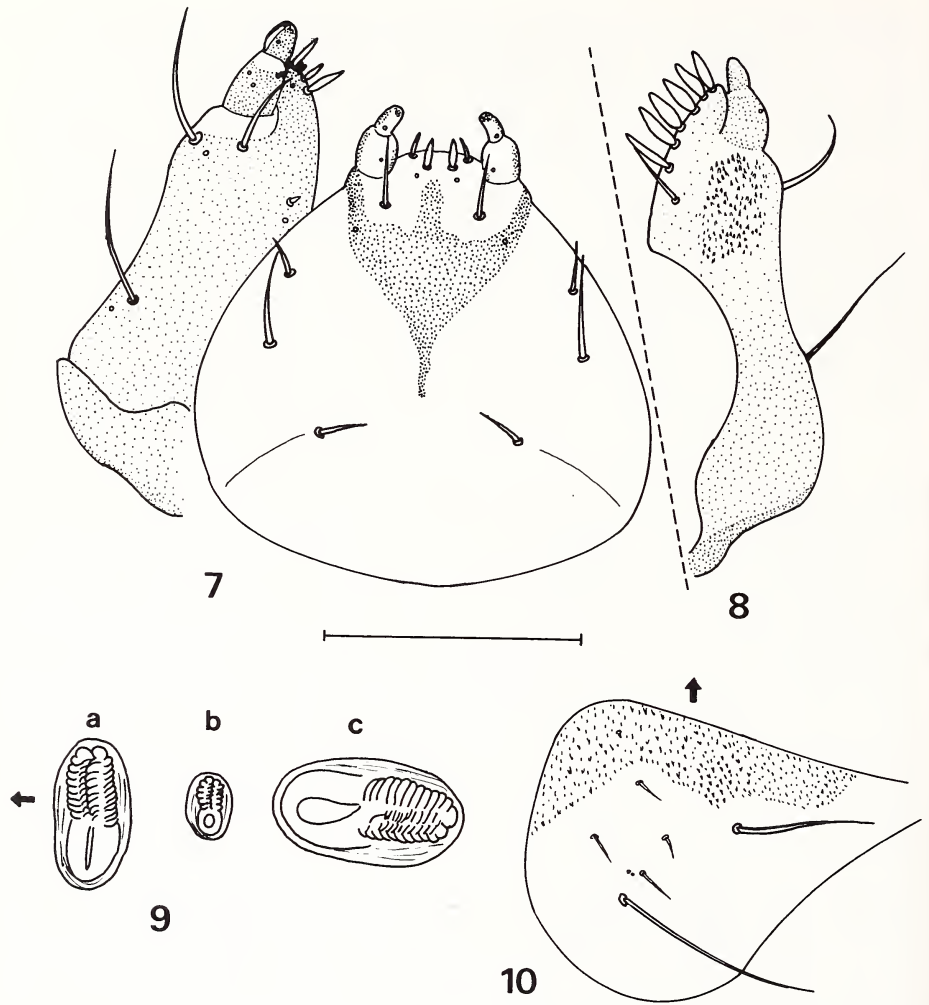
*Head* (Figs. 1, 2) exposed, subcircular, orange-brown pigmented. Frontal line visible in entire length, slightly angulate before apex. Endocarina absent. Postoccipital condyles well developed, obtuse-angled, pigmented. Setae: *fs*4,5 long, subequal, *fs*3 very short, *fs*1,2 minute; *des*1,3,5 long, subequal, *des*2 shorter but well developed, *des*4 very small; *des*3 on epicranium but close to frontal line, *pes* minute; *les*1 short, *les*2 long; *vcs*1,2 short, subequal. Only anterior stemmata distinct, as black pigmented spots. Antennae (Figs. 1, 3) small, set obliquely on anterior margin of head, partially overhanging by frontal projection, antennal sensorium broadly conical. Clypeus (Fig. 4) pigmented at base, with *cls*1,2 well developed. Labrum (Fig. 4) transverse, completely pigmented, the anterior margin slightly projected medianly; *lms*3 well developed but shorter than *lms*1,2; lateral labral sensilla at same level than *lms*1 and as widely separated as *lms*2; labral rods (Fig. 5) strong, joined at base with a distinct stem, Y-shaped. Epipharynx (Fig. 5) with *ams*1 larger as *ams*2 and subequal to *als*; *mes*1 as widely separated as *mes*2; sensillum clusters locate between *mes*1 and *mes*2 and near base of labral rods. Mandibles (Fig. 6) bidentate at apex, with a distinct median projection on cutting edge, *mds*1,2 aligned longitudinally, *mds*1 short, *mds*2 very small, scrobe indistinct. Maxillae (Figs. 7, 8) with 5 *vms* and a row of 7 *dms*; *dms*1 more slender than other *dms*; a patch of minute spinules present below palpus. Labium (Fig. 7) with *pms*2 and *pms*3 pairs separated by same distance, *pms*1 less widely separated; premental sclerite trident-shaped, with subtriangular posterior extension.

*Thorax* (Fig. 11). Spiracle (Fig. 9a) large, with pigmented and finely annulated airtubes, dorsally directed. Pronotum (Fig. 11) with 9 setae plus 2 small epipleural setae. TII, III (Fig. 11) with 1 *prs*; 4 *pds* (*pds*1,2 short, *pds*3,4 long); alar area with 1 *as*; spiracular area with 2 small setae. Pedal area (Fig. 10) with 6 setae (*z* absent) plus minute *x'*; setae *t* and *w* long, others short; 2–5 sensilla present between *v* and *w*.

*Abdomen* (Figs. 11–14). Spiracles (Figs. 9b, c) of different size, with close-set annuli, those of AI–VII (Fig. 9b) smaller than thoracic spiracle, being the interme-

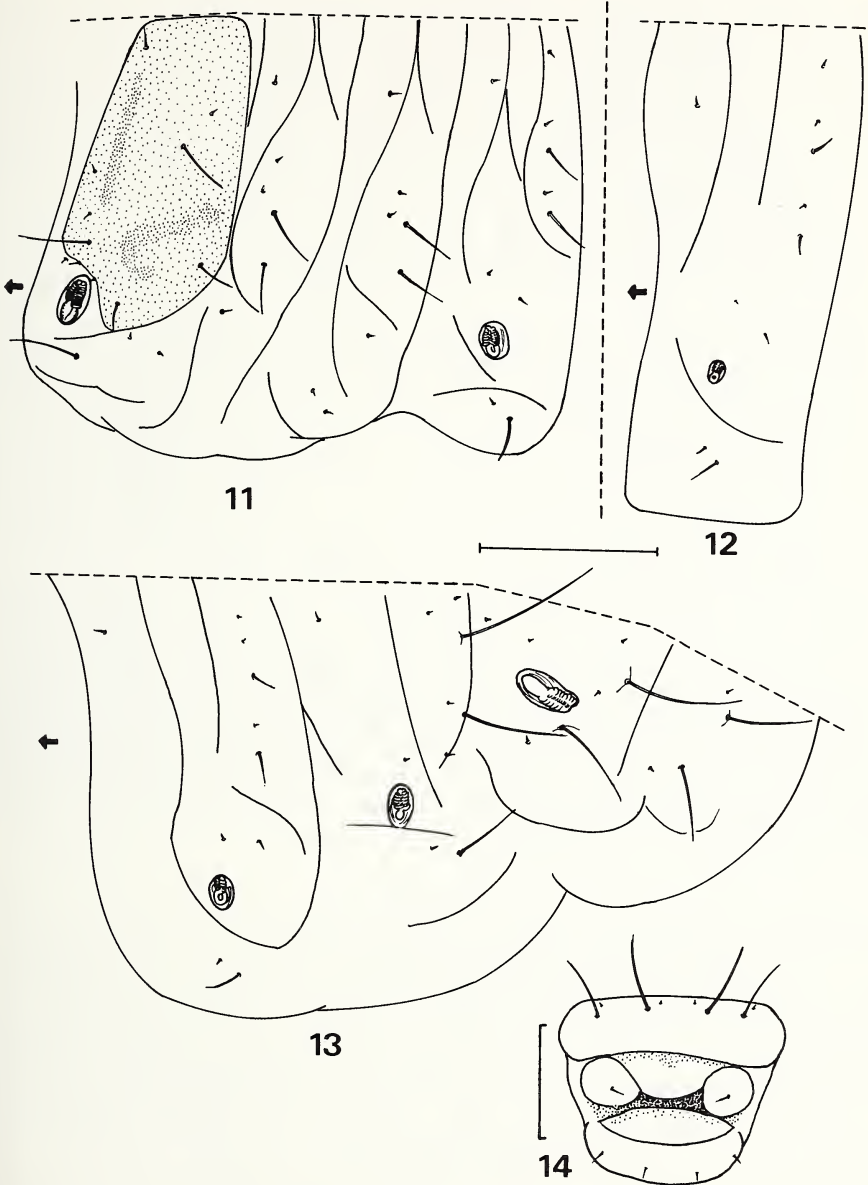


Figs. 1–6. *Rhyparonotus altarensis*, larva. 1, head, dorsal; 2, head, ventral; 3, antenna; 4, clypeus and labrum; 5, epipharynx; 6, mandible. Scales, 1,2 = 1 mm; 3 = 0.1 mm; 4–6 = 0.5 mm.

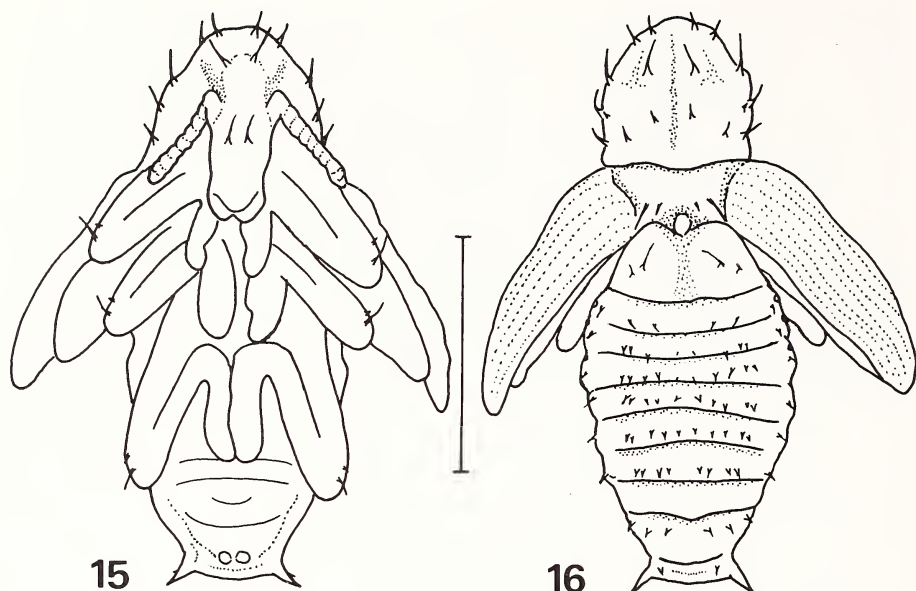


Figs. 7-10. *Rhyparonotus altarensis*, larva. 7, maxilla and labium, ventral; 8, maxilla, dorsal; 9, spiracles (a, thorax; b, AIV; c, AVIII); 10, pedal area. Scales = 0.5 mm.

diate ones smallest, airtubes dorsally directed; AVIII spiracle (Fig. 9c) placed on dorsum and larger than other 7 and than thoracic one, with airtubes posteriorly directed. AI-VII (Figs. 11-13) with *prs* small, vestigial on AVIII; AI-VII with 5 *pds*: *pds*3.5 longer than others, distinctly longer and stronger on AVII; AI-VI with *ss*1.2 on spiracular area; AVII with *ss*2 shifted to postdorsum and *pds*2 set ahead of other *pds*. AVIII (Fig. 13) with 3 *pds* which appear to correspond with *pds*4 (minute), *pds*5 (large and strong) and *ss*2 (minute) of preceding segments. AIX (Figs. 13, 14) with 2 unequal *ds*. AX (Fig. 14) ventral, 4-lobed, anus transversal, lateral lobes smallest bearing 1 small seta.



Figs. 11-14. *Rhyparonotus altarensis*, larva. 11-13, one side, dorsolateral; 11, TI-III, AI; 12, AIV; 13, AVI-X. 14, AIX, X, caudal view. Scales = 1 mm.



Figs. 15, 16. *Rhyparonotus altarensis*, pupa, habitus. 15, ventral; 16, dorsal. Scale = 5 mm.

*Pupa* (Figs. 15, 16). Maximum length 11.95 mm. Setae as in Figs. 15, 16. Cuticle glabrous. Setae orange-brown, tapering, moderately strong on head and pronotum, smaller on abdomen, mounted on small tubercles. Antennal club smooth. Each femur with a pair of unequal setae. Secondary pterotheca shorter than primary. Spiracles distinct, with dark peritreme. AIX with prominent posterior processes, each with a small associated seta located laterally at base.

#### DISCUSSION

The larva of *Rhyparonotus altarensis* presents some characters that are similar to those of *Anchonus* spp. indet. studied by Anderson (1952), such as: endocarina absent, antennae overhung by frontal projection, labrum completely pigmented with anterior margin produced medianly, maxillary mala with 7 *dms*, AI-VII (typical abdominal segments) with 3 dorsal folds, spiracle on AVIII dorsal and larger than those of anterior segments, and body asperities fine. These features may be useful to diagnose the tribe in the larval stage, but they are also present among other molytine larvae. The anchonine larvae herein studied and those examined by Anderson (1952) are different, however, in characters involving the position of *des3*, shape of labrum and position of lateral sensilla, shape of labral rods, arrangement of *als* on epipharynx, structure of mandible on cutting edge (inner margin), orientation of spiracular airtubes on AI-VII, relative length of *pds5* on AI-VII, and number of *pds* on AVIII and AIX. These characters may be diagnostic at generic level within Anchonini.

According to May (1993), Molytinae comprise two distinct elements: "live tissue feeders" (=tribes Molytini and Hylobiini), with modal mouthparts, and "inhabitants

of dead wood" (=tribe Phrynixini), with mouthparts adapted to xylophagy. The larvae of *Rhyparonotus* herein studied, apparently subcortical borders in dying trunks, fall in the former group, characterized by the pigmented head, transverse labrum, clear hypopharyngeal bracon, broadly conical antennae, and maxilla with 2 setae on palpifer and 7 *dms*, plus TII, III with long *pds4*. These characters, however, appear to be plesiomorphic when compared with other weevil larvae, and thus, provide no evidence for monophyly. The latter group may correspond instead with a monophyletic group, because although several mouthpart characters defining it are hypothesized as "adaptations for xylophagy" (May, 1993), their maxillae with 3 setae on palpifer and 10 *dms*, and the short *pds4* on TII, III may be apomorphies of *Phrynixus* and its allies.

May (1993) considers "labral rods separate, subparallel or bowed" to diagnose Molytinae (from the Australian Region). However, larvae of *Rhyparonotus* herein studied, as well as those of *Heilipodus erythropus* (Rosado-Neto, 1980), belonging to the tribe Hylobiini, show Y-shaped labral rods (joined at base with a distinct stem), which could be phylogenetically informative. It is worth noting that, among other Curculionidae, labral rods Y-shaped define the larvae of Cryptorhynchinae, which can be distinguished from those of Molytinae by their lateral AVIII spiracles (May, 1993, 1994). We have noted that the 3 *pds* on AVIII of *Rhyparonotus* larvae (and probably other molytines, as suggested by figures in literature) appear to correspond with *pds4*, *pds5* and *ss2* of preceding segments. This is different in larvae of Rhytirrhini and some Aterpini (Cyclominae), that lack *pds5* and *ss2* on AVIII (Marvaldi, 1998). Although there are examples of molytine larvae with *des3* on frontal line or on frons (e.g., Anderson, 1952:291; May, 1993:82), those of typical Molytinae (e.g., *Hylobius*) have *des3* on epicranium. The latter character allows separation from the morphologically similar larvae of Aterpini with *des3* always on frons or on frontal line (May, 1994). To this could be added that most (though not all) larvae of Molytinae have only 2 *ds* on AIX. All these characters also provide separation from larvae of Rhytirrhini, which in addition are diagnosed by their very reduced lateral labral setae. Larvae of Molytinae, with distinct postoccipital condyles and AVIII spiracles on dorsum, can be separated quite easily from those of Cossoninae and Scolytinae (Marvaldi, 1997), but they are difficult to distinguish, morphologically, from some Curculioninae *s. str.* (May, 1993, 1994).

Further studies on immature stages, including some less "typical" Molytinae, e.g., Petalochilini or Juanorhinini, are still needed in order to test the monophyly of Molytinae. Similarly, a more complete sample of larvae representative of Anchonini would be valuable to define this tribe at the larval stage and to ascertain the phylogenetic relationships of the Anchonini.

#### ACKNOWLEDGMENTS

We are indebted to Jane Beard of the Natural History Museum, London, for making available literature to us. This study was supported by the "Consejo Nacional de Investigaciones Científicas y Técnicas" (CONICET) and by a grant of "Fundación Antorchas," Argentina.

#### LITERATURE CITED

- Anderson, D. M. 1991. Curculionidae (broad-sense) (Curculionoidea). Pp. 594–612. In: Stehr, F. W. (ed.), *Immature Insects*, Vol. 2. Kendall/Hunt Publishing Company.

- Anderson, W. H. 1947. A terminology for the anatomical characters useful in the taxonomy of weevil larvae. *Proc. Ent. Soc. Washington* 49:123-132.
- Anderson, W. H. 1952. Larvae of some genera of Cossoninae (Coleoptera: Curculionidae). *Ann. Entomol. Soc. Amer.* 45(2):281-309.
- Böving, A. G. and F. C. Craighead. 1930. An illustrated synopsis of the principal larval forms of the order Coleoptera. *Entomol. Amer.* 11(1-4):1-351.
- Calder, 1989. The alimentary canal and nervous system of Curculionoidea (Coleoptera): Gross morphology and systematic significance. *J. Nat. Hist.* 23:205-265.
- Faust, J. 1892. Die Anthoniden-Gruppe. *Dtsch. Ent. Zeitschr.* 1892:17-60.
- Gardner, J. C. M. 1933. The early stages of two Indian weevils. *Stylops* 2:81-85.
- Gardner, J. C. M. 1934. Immature stages of Indian Coleoptera (14) (Curculionidae). *Indian Forest Records* 20(2):1-42.
- Gardner, J. C. M. 1938. Immature stages of Indian Coleoptera (24 Curculionidae Contd.). *Indian Forest Records, Entomol. Ser.* 3(12):227-261.
- Kuschel, G. 1987. The subfamily Molytinae (Coleoptera: Curculionidae): General notes and descriptions of new taxa from New Zealand and Chile. *New Zealand Entomol.* 9:11-29.
- Kuschel, G. 1995. A phylogenetic classification of Curculionoidea to families and subfamilies. *Memoir Entomol. Soc. Wash.* 14:5-33.
- Lee, C. and K. Morimoto. 1988. Larvae of the Weevil Family Curculionidae of Japan. Part 2. Hyperinae to Cioninae (Insecta: Coleoptera). *J. Fac. Agr., Kyushu Univ.* 33(1-2):131-152.
- Marshall, G. A. K. 1932. Notes on the Hylobiinae (Col., Curc.). *Ann. Mag. Nat. Hist. ser. 10,* 9:341-355.
- Marvaldi, A. E. 1997. Higher level phylogeny of Curculionidae (Coleoptera: Curculionoidea) based mainly on larval characters, with special reference to broad-nosed weevils. *Cladistics* 13:285-312.
- Marvaldi, A. E. 1998. Larvae of South American Rhytirrhinae (Coleoptera: Curculionidae). *Coleopt. Bull.* 52(1):71-89.
- May, B. M. 1971. Entomology of the Aucklands and other islands south of New Zealand: Immature stages of Curculionoidea. *Pacific Insects Monograph* 27:271-316.
- May, B. M. 1973. A new species of *Megacolabus* and descriptions of the immature stages of *M. decipiens* (Coleoptera: Curculionidae). *Journal of the Royal Society of N. Z.* 3:255-262.
- May, B. M. 1979. Immature stages of Curculionidae: the "rat-tailed" larva and the pupa of *Notonesius aucklandicus* (Phrynixinae). *N. Z. Journal of Zoology* 6:577-582.
- May, B. M. 1981. Immature stages of Curculionoidea—the weevils of The Snares Islands, New Zealand. *N. Z. Journal of Zoology* 8:255-280.
- May, B. M. 1987. Immature stages of Curculionidae (Coleoptera): the larva and pupa of *Karocolens pittospori* (Molytinae). *N. Z. Entomologist* 9:29-34.
- May, B. M. 1993. Larvae of Curculionoidea (Insecta: Coleoptera): A systematic overview. *Fauna of New Zealand* 28:1-223.
- May, B. M. 1994. An introduction to the immature stages of Australian Curculionoidea. Pp. 365-755. *In: Zimmerman, E. C., Australian Weevils, Vol. II.* Melbourne, CSIRO.
- Morimoto, K. 1982. The family Curculionidae of Japan. I. Subfamily Hylobiinae. *Esakia* 19: 51-121.
- Morrone, J. J. 1997. Argentinian weevils (Coleoptera: Curculionoidea): Preliminary overview, with nomenclatural and distributional notes. *Physis* (Buenos Aires) (in press).
- Morrone, J. J. and S. A. Roig-Juñent. 1995. The diversity of Patagonian weevils: An illustrated checklist of the Patagonian Curculionoidea (Insecta: Coleoptera). *L. O. L.A., Buenos Aires*, 189 pp.

- Oberprieler, R. G. 1988. The life history of *Paramecops stapeliae* (Marshall), with a review of the genus *Paramecops* (Coleoptera: Curculionidae: Molytinae). J. Nat. Hist. 22:1451–1464.
- O'Brien, C. W. and G. J. Wibmer. 1982. Annotated checklist of the weevils (Curculionidae *sensu lato*) of North America, Central America, and the West Indies (Coleoptera: Curculionoidea). Mem. Amer. Ent. Inst. 34:382 pp.
- Peterson, 1951. Larvae of insects. Part II Coleoptera, Diptera, Neuroptera, Siphonaptera, Mecoptera, Trichoptera. 1st ed., Edward Bros., Inc., Ann Arbor, 134 pp.
- Rosado-Neto, G. H. 1980. Description of larva and pupa of *Heilipodus erythropus* (Klug, 1829) (Coleoptera, Curculionidae). Rev. bras. Ent. 24(2):111–115.
- Scherf, H. 1964. Die entwicklungsstadien der mitteleuropäischen Curculioniden (Morphologie, Bionomie, Ökologie). Abh. Senckenberg. Naturf. Ges. 506:1–355.
- Thompson, R. T. 1992. Observations on the morphology and classification of weevils (Coleoptera, Curculionoidea) with a key to major groups. J. Nat. Hist. 26:835–891.
- Voisin, J.-F. 1992. *Gibbanchonus* n. gen. et *G. krausei* n. sp. du Mato Grosso (Brésil) (Insecta, Coleoptera, Curculionidae: Molytinae). Reichenbachia Mus. Tierk. Dresden 29(18):93–95.
- Voisin, J.-F. 1993. Notes sur la tribu des Anchonini: 2. Description de quatre genres nouveaux et revue de six anciens (Coleoptera, Curculionidae). Nouv. Rev. Ent. (N.S.) 10(4):327–340.
- Wibmer, G. J. and C. W. O'Brien. 1986. Annotated checklist of the weevils (Curculionidae *sensu lato*) of South America (Coleoptera: Curculionoidea). Mem. Am. Entomol. Inst. (39):i–xvi, 1–563.
- Zimmerman, E. C. 1994. Australian Weevils, Vol. I. Melbourne, CSIRO, 741 pp.

Received 30 January 1998, accepted 18 August 1998.

**Appendix.** Tribes assigned to Molytinae (after Kuschel, 1987; Thompson, 1992; Morrone, 1997). Genera between brackets are those for which larvae have been described.

Acicnemidini Lacordaire

Amalactini Lacordaire

Anchonini Imhoff (*Anchonus* [Anderson, 1952]).

Bagoini Thomson (*Bagous* [Scherf, 1964; May, 1994]).

Cholini Schoenherr

Cycloterini Marshall

Dinomorphini Lacordaire

Emphyastini Lacordaire

Euderini Lacordaire

Galloisiini Morimoto

Guioperini Lacordaire

Haplonychini Lacordaire

Hylobiini Kirby (=Heilipinae Faust) (*Acles* [Gardner, 1938]; *Heilipodus* [Rosado-Neto, 1980]; *Heilipus* [Böving & Craighead, 1930]; *Hylobius* [Gardner, 1938; Scherf, 1964; Anderson, 1991]; *Kobuzo* [Gardner, 1938]; *Pagiophloeus* [Gardner, 1934]; *Paramecops* [Gardner, 1934; Oberprieler, 1988]).

Ithyporini Lacordaire (=Conotrachelini) (*Conotrachelus* [Peterson, 1951; Anderson, 1991]).

Juanorhinini Aurivillius

Lepyrini Faust

Lithinini Lacordaire

- Lymanini Lacordaire (genus indet. near *Ithaura* [Anderson, 1952]).
- Mecysolobini Reitter (*Mesalcidodes* [Lee & Morimoto, 1988]).
- Molytini Schoenherr (=Liparini Latreille) (*Arecophaga* [May, 1993]; *Hadramphus* [May, 1971, 1981]; *Karocolens* [May, 1987]; *Liparus* [Scherf, 1964]; *Lyperobius* [May, 1981]; *Paedaretus* [May, 1993]).
- Nettarhinini Lacordaire
- Omophorini Marshall
- Pacholenini Lacordaire
- Paipalesomini Marshall (*Paipalesomus*, *Peribleptus* [Gardner, 1993]).
- Petalochilini Lacordaire
- Phoenicobatini Champion
- Phrynixini Kuschel (*Megacolabus* [May, 1973]; *Notonesius* [May, 1979]; *Phrynixus* [May, 1971, 1981]; *Rhysheus* [May, 1993]).
- Pissodini Thomson (*Pissodes* [Anderson, 1947; Scherf, 1964]).
- Sternechini Lacordaire (*Chalcodermus* [Peterson, 1951; Anderson, 1991]).
- Styanacini Chujo & Voss
- Trypetidini Lacordaire (*Heteramphus*, *Nesotocus* [Anderson, 1952]).
- Molytinae *insertae sedis* (*Demysus*, *Melanotrane*s, *Orthorhinus*, *Tranes* [May, 1994]).